

# Resolution of fossil monocotyledonous roots to the natural taxa

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## ABSTRACT

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Monocotyledons possess endogenously originated adventitious roots. Such permineralized roots occur abundantly in the Cretaceous and onward horizons assigned to the artificial genera *Rhizopalmoxylon* Gothan, *Aerorhizos* Chitale, *Velamenorhizos* Barlinge and Paradkar and *Hygrorhizos* Trivedi *et al.*. Fossil roots reported so far have not been described with uniform terminologies which lack precision and delimitations of the tissues. Considering a need to have uniform terminologies in the description of permineralized roots, a system based upon Seubert's work has been proposed to describe the monocotyledonous roots.

**Key-words**—Monocotyledons, Permineralization, Anatomy, Roots.

प्राकृतिक टैक्सा के लिए एकबीजपत्री जड़ों के जीवाश्मों का वियोजन

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सारांश

एकबीजपत्री अंतःप्रवर्धी रूप से उत्पन्न अपस्थानिक जड़ों को नियंत्रण में रखती है। ऐसी पर्मिनीकृत जड़ें कृत्रिम वंश *राइज़ोपामॉक्सिलॉन* गोथान, *एरोराइज़ॉस* चिताले, *वेलामेनोराइज़ॉस* बर्लिंगे एवं पराडकर तथा *हाइग्रोराइज़ॉस* त्रिवेदी एवं अन्य को निर्धारित क्रिटेशस एवं अग्रवर्ती संस्तर-स्थितियों में प्रचुरता से प्राप्त होती हैं। अब तक प्राप्त जीवाश्म जड़ें एक रूप शब्दावलियों से वर्णित नहीं हुई हैं जो कि ऊतकों की परिशुद्धता एवं परिसीमन का अभाव है। पर्मिनीकृत जड़ों के वर्णन में एक रूप शब्दावलियों की आवश्यकता पर विचार करते हुए, एकबीजपत्री जड़ों को वर्णित करने के लिए स्युबर्ट के शोध कार्य पर आधारित एक पद्धति प्रस्तावित है।

**संकेत-शब्द**—एकबीजपत्री, पर्मिनीकरण, शारीर, जड़ें।

## INTRODUCTION

Monocotyledons constitute about 59,300 species assigned to 112 families comprising about 25 % of the angiosperms (Thorne, 2002). Anatomy of the

monocotyledons has been investigated by large number of workers. However, their roots except those of palms have been poorly addressed (Haberlandt, 1914; D'Almeida & Ramaswamy, 1948; Surange, 1950; Fahn, 1954; Metcalfe, 1960, 1961, 1963, 1971; Cutler,

1969; Tomlinson, 1969, 1970, 1982; Ayensu, 1972; Ancibor, 1979; Tomlinson & Wilder, 1984; Keating, 2003).

### ROOT ANATOMY OF EXTANT PALMS

Anatomy of roots in palms has been investigated by many workers. Mohl (1845, 1849) was the pioneer to describe the characteristic arrangement of the vascular tissues and their connections with the stem vascular bundles. He has also noted the presence of characteristic medullary bundles that occur in palms only. Karsten (1847) noted a single undifferentiated apical meristem. Naegeli (1858), Russow (1875), Falkenberg (1876), De Bary (1877), van Tieghem (1870), Olivier (1880) and Mangin (1882) made further additions to the root anatomy. Cormack (1896) noted polystelic condition in number of palms like - *Areca*, *Caryota*, *Corypha*, *Dypsis*, *Euterpe*, *Geonoma*, *Hyophorbe*, *Iriarteia*, *Kentia*, *Livistona*, *Phoenix*, *Ptychosperma* and *Verschaffeltia*. Gillain (1900) noted three meristematic layers in the root apex of *Hydriastele* (*Kentia*) and analyzed the root structure in number of palms. Drabble (1904) investigated 67 species. Mahabale and Udawadia (1960) analyzed 37 adult palm roots and initiated the work on the resolution of the fossil organ genus *Rhizopalmoxyton* to the natural taxa. Tomlinson (1961, 1990) has studied the structural biology of palms along with the roots. Seubert (1996a, b, 1997, 1998a, b) made in-depth anatomical analysis of 159 genera based upon the ontogenetical

and phylogenetical considerations adopted by Guttenberg (1960, 1968) and Hanstein (1870) and suggested the usage of a system of uniform terminologies in describing extant palm root.

### PERMINERALIZED MONOCOTYLEDONOUS ROOTS

Monocotyledonous adventitious roots have been described from the Cretaceous and onward horizons. Majority of them are preserved due to permineralization, the most informative mode of preservation for the study of fossilized plants. In this process infiltration and permeation of the tissues occur by mineral charged water. Over a period of time, intracellular and interstitial precipitation of the dissolved mineral results in hardening within the plant and it effectively becomes solid rock. Thus, although the plant cell walls still consist of organic matter, they have been chemically altered and the intercellular spaces and cell lumens are filled with precipitated minerals (Schopf, 1975). The characters of the roots are ought to be present in them and the root anatomical details are available. It would be better to circumscribe the fossil forms into their nearest possible natural taxa.

The permineralized monocotyledonous roots are mainly assigned to the artificial organ genus *Rhizopalmoxyton* Gothan assuming their affinities with the palms (Family–Arecaceae). However, these could belong to the other monocotyledons also. Felix (1883) reported the institution of the monocotyledonous organ

#### PLATE 1

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| <ol style="list-style-type: none"> <li>1. Schematic transverse section (T.S.) of a palm root showing different tissues.</li> <li>2. T.S. of root of <i>Calamus</i> sp.</li> <li>3. <i>Phoenix reclinata</i> Jacq.</li> <li>4. <i>Bentinckia nicobarica</i> Becc.</li> <li>5. <i>Daemonorops</i> sp.</li> <li>6. <i>Caryota mitis</i> Lour.</li> <li>7. <i>Phytelephas</i> sp.</li> <li>8. <i>Raphia australis</i> Oberm. &amp; Strey</li> <li>9. <i>Copernicia macroglossa</i> H. Wendl. ex Becc.</li> <li>10. <i>Pritchardia beccariana</i> Rock</li> <li>11. <i>Paspalum commessonii</i> Lam. (Gramineae) : a. Entire cross section, b. part enlarged showing very large</li> </ol> | <div style="text-align: right; margin-bottom: 10px;">→</div> <ol style="list-style-type: none"> <li>intercellular spaces, thick walled pericycle and reduced vascular tissue</li> <li>12. <i>Triuris</i> sp. (Family: Triuridaceae : Helobiae)</li> <li>13. <i>Smilax herbacea</i> Linn.: a, Entire transverse section, b. part enlarged showing multilayered pericycle</li> </ol> <p>r – rhizodermis, v- velamen, e– exodermis, c – cork, oc – outer cortex, ic– inner cortex, en–endodermis, vc – vascular cylinder, fb – fibre bundle, f – fibre, m–mucilage cell, scl–sclerenchyma, I.C.–intercellular cavity, C.P.–cortical parenchyma, E.P.–epidermis, END–endodermis.</p> <p>1-10. after Seubert, 1996, 1997, 1998; 11. after Metcalfe, 1960; 12. after Tomlinson, 1982 &amp; 13. after Jeffrey, 1917.</p> |
|---|---|

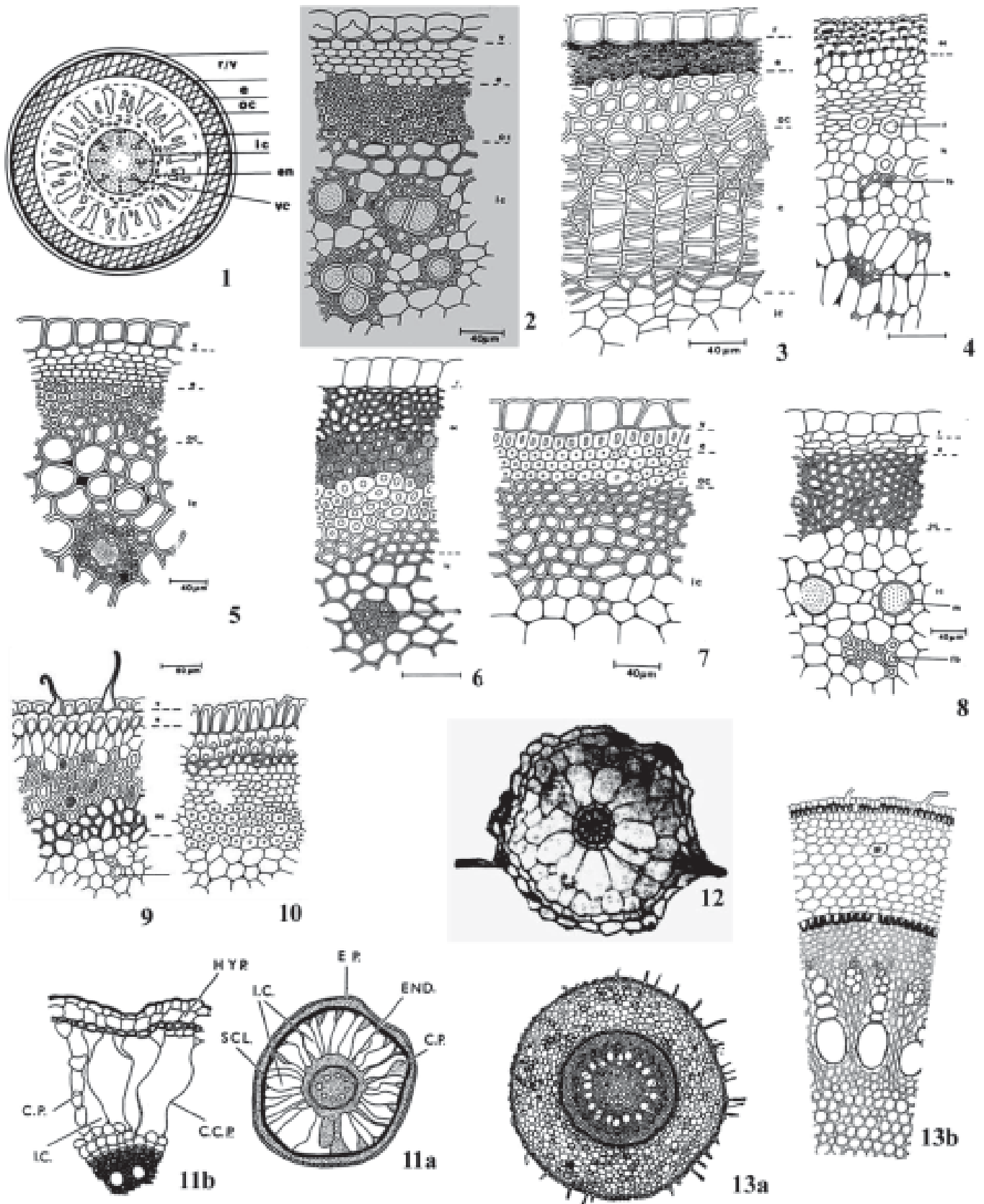


PLATE 1

genus for permineralized roots as *Rhizopalmoxyton* showing their affinities with palms. However, he did not provide the description or diagnosis and as such *Rhizopalmoxyton* Felix (*loc. cit.*) remained as *Nomen nudum*. Establishment of *Rhizopalmoxyton* became valid when Gothan (1942) provided its diagnosis and description. *Rhizopalmoxyton* Gothan possesses 13 species, viz., *R. glaseli* Gothan (1942), *R. bohlenianum* Gothan (1942), *R. libycum* Koeniguer (1970), *R. behuninii* Tidwell *et al.* (1972), *R. blackii* Tidwell *et al.* (1972), *R. scottii* Tidwell *et al.* (1972), *R. sundaram* Mahabale and Rao (1973), *R. huepaciense* Cevallos-Ferriz and Ricalde-Moreno (1995), *R. teguachiense* Cevallos-Ferriz and Ricalde-Moreno (1995), *R. borassoides* Awasthi *et al.* (1996), *R. angiorhizon* (= *Palmoxyton angiorhizon* Stenzel, 1904; Bonde *et al.* 2009), *R. macrorhizon* (= *Palmoxyton macrorhizon* Stenzel, 1904; Bonde *et al.* 2009) and *R. singulare* Bonde *et al.* (2009). In addition, a root comparable to *Nypa* from the Mohgaonkalan Intertrappean beds (Verma, 1974) and a borassoid root from Nawargaon Intertrappean sediments (Ambwani, 1981) have been reported.

Some of the fossil palm stems like *Palmoxyton rutherfordi* Stockmans & Williere (1943), *Palmoxyton* sp. Stockmans & Williere (1943), *P. sclerodermum* Sahni (Shukla, 1946), *P. surangei* Lakhanpal (1955), *P. kraeuselii* Rao & Menon (1966), *P. raoi* Menon (1968), *P. macginitie* Tidwell *et al.* (1971) and *P. sundaram* Sahni (Bonde *et al.*, 2004) possess basal regions attached with the roots. In addition, roots of a scitaminean plant *Cyclanthodendron sahnii* Sahni and Surange (Sahni & Surange, 1953; Biradar & Bonde, 1990); *Sparganium* sp. (Mahabale, 1953); a cyperaceous rhizome *Scirpusoxyton indicum* Shete (1989); a viny araceous axis *Rhodopathodendron tomlinsonii* Bonde (2000); a pandanaceous rhizome *Pandanusoxyton kulkarnii* Patil and Datar (2002) and a liliaceous corm *Eriosphermocormus indicus* Bonde (2005) have been described. In the light of the present system, reexamination of roots of these taxa will throw more light on the affinities with the natural taxa.

*Hygrorhizos deccanii* Trivedi *et al.* (1985) is a root comparable to *Hygrorhiza* (Family – Poaceae). *Velamenorhizos intertrappeanum* Barlinge and Paradkar (1978) is an aerial abortive root with velamen showing its affinities with Araceae, Arecaceae and Orchidaceae whereas *Aerorhizos harrisii* Chitaley (1968) is an aquatic root showing its resemblance with *Alpinia*, *Canna*, *Cyperus*, *Hedychium* and other aquatic plants. A doubtful root of *Eichhornia* (Family – Pontederiaceae) has also been reported (Patil & Singh, 1978; Awasthi *et al.*, 1996).

### SYSTEM FOR RESOLUTION OF FOSSIL MONOCOTYLEDONOUS ROOTS

The fossil monocotyledonous roots reported so far have not been described with uniform anatomical terminologies. They lack precision and delimitations of the tissues particularly those of the extra stelar region. A system based upon Seubert's anatomical work on extant palms has been proposed to resolve the fossil monocotyledonous roots including the palms to their natural taxa (Seubert, 1996a, b, 1997, 1998a, b). It may throw some light on the phylogeny of monocotyledons in future (Pl. 1.1-13).

Monocotyledonous roots exhibit the following tissues from periphery to the centre. A. *Extrastelar region* - Rhizodermis / Velamen; Exodermis; Outer cortex; Inner cortex; and Endodermis. B. *Stelar region* – Pericycle; Sheathing layer of vascular tissue; Xylem and Phloem bundles; Pith and Medullary bundles.

*Rhizodermis/Velamen*—*Appendages* - Root spines – present/absent, pointed/blunt; root tubercles-spherical/bottle like; pneumathodes - present/absent; root hairs present/absent; lateral roots –few/dense, elongate/cylindrical; branching system - fewer, short/long, thick/thin.

*Rhizodermis*—Cells hexagonal/tangentially elongated/honey combed, expanded in either direction/irregularly expanded; cell wall thickened/unthickened.

*Velamen*—one/many layered; cell wall equally/moderately/strongly/unequally thickened, outer walls thicker than inner walls; root hairs present/absent.

*Exodermis*—Thickness one/many layered; cell walls moderately/strongly thickened; tannin contents present/absent.

*Outer cortex*—a. *Three zoned*, middle zone different from outer and inner parenchymatous zones; pits on cell wall present/absent; raphides/tannin cells present/absent, isolated/in files; cell thickness unthickened/strongly thickened.

b. *Two zoned*, outer zone with thin walled cells, inner zone with thick walled cells.

c. *Homogeneous*.

d. Rhizodermis, exodermis, outer cortex absent/cork present.

*Inner cortex*—Homogeneous/three zoned; aerenchyma absent/weakly/strongly developed; air spaces small/large/quadrangular/radially elongated; very large in one/many rings; stone cells present/absent, solitary/forming hollow cylinder; fibres – present/absent; outline circular/irregular; solitary/forming bundles; fibre bundles – *Kentia* type/*Raphia* type .

*Endodermis*—Layers-one/many; cell wall unthickened/equally thickened/unequally thickened/‘U’ shaped thickened; casparian thickening present/absent.

*Vascular cylinder*—Eustelic/polystelic .

*Pericycle*—Layers one/several; fibre cells present/absent.

*Sheathing layer*—Sclerotic/parenchymatic.

*Xylem / Phloem bundles*—Size, shape; number.

*Pith*—Parenchymatic/sclerenchymatic.

*Medullary bundles*—Present/absent; number.

This system may prove to be the most feasible method to resolve the permineralized monocotyledonous roots to the extant taxa which may throw light on their phylogeny.

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