# Life and time of Indian Williamsonia

Jayasri Banerji

Banerji, J 1992. Life and time of Indian Williamsonia. Palaeobotanist 40: 245-259.

The Williamsonia plant, belonging to the order Bennettitales, consists of stem—Bucklandia Presl, leaf— Ptilophyllum Morris, male flower—Weltrichia Braun and female flower—Williamsonia Carruthers. This plant was perhaps a small, much branched woody tree of xerophytic environment. It co-existed alongwith extremely variable and rich flora including highly diversified plant groups from algae to gymnosperms. In India, it appeared during the marine Jurassic, proliferated and widely distributed in the Lower Cretaceous and disappeared from the vegetational scenario of Upper Cretaceous Period with the advent of angiosperms.

Key-words-Bennettitales, Williamsonia, Jurassic-Cretaceous (India).

Jayasri Banerji, Birbal Sabni Institute of Palaeobotany, 53 University Road, Lucknow 226 007, India.

## सारौंश

#### भारतीय विलियमसोनिया का जीवनकाल

#### जयश्री बैनर्जी

बैन्नेटाइटेलिस क्रम से सम्बद्ध विलियमसोनिया पौधे के विभिन्न अवयव अलग-अलग नामों से जाने जाते हैं इसका तना—बुक्लैंडिया प्रेस्ल, पत्ती—टाइलोफिल्लम् मोरिस, नर पुष्प—बेल्ट्राइकिया ब्रॉन एवं मादा पुष्प—विलियमसोनिया कैरुथर्स से नामॉकित किया जाता है। यह पौधा शुष्क वातावरण में उगने वाला एक छोटा, अधिक शाखाओं से युक्त वृक्ष था। यह विभिन्नता से युक्त वनस्पतिजात, जिसमें शैवालों से लेकर अनावृतबीजी समूह के पौधे विद्यमान थे, का ही एक अवयव है। इसका समुद्री जूराई कल्प में उदय हुआ, अधरि क्रीटेशी कल्प में विकसित एवं दूर-दूर तक फैला तथा उपरि क्रीटेशी कल्प में आवतबीजी पौधों के उदय होने के साथ-साथ विलप्त हो गया।

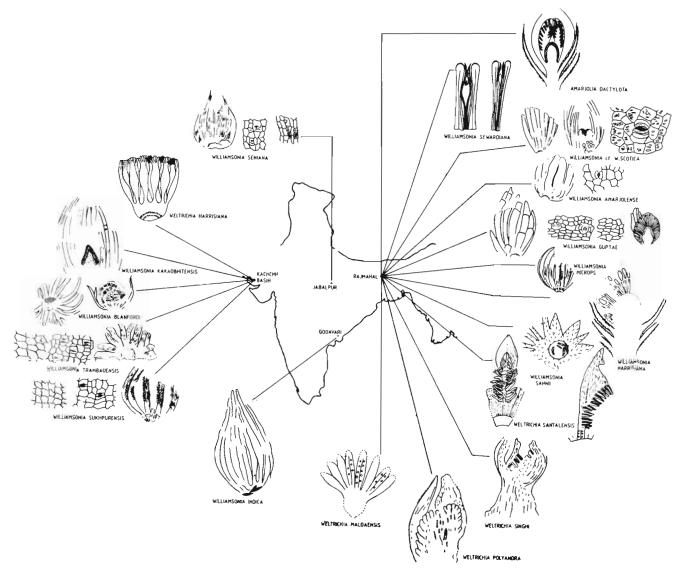
## LIFE OF WILLIAMSONIA PLANT

In the Upper Mesozoic Era, a new group of plants—the Bennettitales, developed that colonised various habitats of terrestrial areas. This group continued till Early Cretaceous. In India, it is represented by the members of the family Williamsoniaceae and occurs pre-dominantly during Lower Cretaceous Period (Text-figure 1). The *Williamsonia* plant is known through its stem— *Bucklandia* Presl, leaf—*Ptilophyllum* Morris, male flower—*Weltrichia* Braun, and *Williamsonia* Carruthers female flower.

## Stem-Bucklandia Presl

The *Bucklandia* stem was erect and cylindrical, protected by armour of rhomboidal leaf-bases. Four species of *Bucklandia* described from India are: *B. indica* Seward, *B. sabnii* Bose, *B. guptai* Sharma and

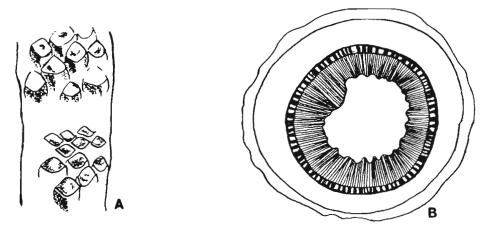
B. dichotoma Sharma. In B. indica Seward, the secondary wood is more compact than recent cycads and cycadeoids. The medullary rays are uniseriate and tracheids have multiseriate bordered pits on their radial walls instead of scalariform pitting in the majority of cycadeoid stems. Secretory canals are abundant in the parenchymatous ground tissue, the cambium and phloem are not preserved. The cross section (Text-figure 2) of B. sahnii Bose 1953a shows well-defined growth rings in the secondary wood, 4-9 cell thick cork cambium present outside the cortex. Cortex is fairly wide, parenchymatous with thick-walled cells, singly or in group and leaf traces with a single bundle which finally breaks up into 5-7 strands. Pericycle is with few layers of polygonal cells. The phloem ring is marked by alternate bands of poorly developed rays leaving big



Text-figure 1-Distribution of Indian Williamsonian flowers.

cavities. Phloem consists of alternate thick- and thinwalled elements. Traces of cambium are seen below the phloem. The tracheids of early wood have a large diameter with a thinner wall than those of the late wood. The late wood is remarkably developed and occupies about two-third of the whole growth rings. Radial surface of the late wood tracheids have not so variable pitting but the early wood shows different type of pitting from scalariform to multiseriate. Ray cells in radial view are rectangular and much wider than high. The pits in the field are bordered, usually one to six or more, oval or circular in shape. Longitudinal section shows that the inner-most part of xylem ring consists of 4 to 8 layers of slender tracheids with spiral thickening in their walls. Pith is very wide and consists of parenchymatous cells with numerous scattered thick-walled cells similar to the cortex. In *B. sabnii* the stem had forked branches and the flowers were borne at the point of bifurcation as in *Wielandiella angustifolia* Nathorst. *Bucklandia guptai* Sharma 1967 has sparsely arranged symmetrical leaf basis and anatomically it shows the presence of numerous growth rings and mostly uniseriate medullary rays which are rarely biseriate. *B. dichotoma* Sharma 1969b is characterised by having longer rays, biseriate in nature and bordered pits with narrow elliptical pit pores. According to Bose (1974, p. 196), this species is based on variable characters and these should not be used for specific differentiation. Besides, he considers it perhaps as a young stem.

*Mode of branching*—Seward (1912), while describing petrified *Williamsonia* from Scotland, suggested the existence of a sympodial branching,



Text-figure 2-A, Bucklandia-stem; B, T.S. of Bucklandia-stem.

while Sahni (1932) suggested monopodial branching in *W. sewardiana*.

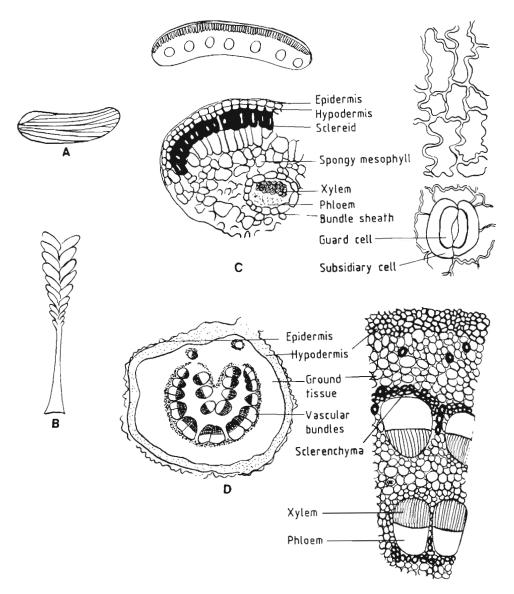
#### Leaf-Ptilopbyllum Morris

The genus *Ptilophyllum* Morris in Grant (1840) originally comprised pinnate fronds, pinnae with acute or obtuse apices. The circumscription of the genus was later enlarged to include pinnate frond, pinnae linear falcate in shape with acute, sub-acute to obtuse apices, basal acroscopic margin round, basal basiscopic margin decurrent and veins subparallel to slightly diverging towards the apical margin. The rachis contains a double arc of collateral vascular bundles comparable to the pteridosperms (Text-figure 3). Sclerenchymatous patches and sclereids are found in the cortex. Single bundle arises from the upper corner of the double arc of vascular bundle of rachis which divides into two at the base of the pinna. It is repeatedly forked at different levels to form a more or less parallel venations.

The anatomy of pinna shows collateral, exarch xylem with a parenchymatous bundle sheath. Sclereids occur isolated or in groups in the palisade and spongy mesophyll. The salient features of the cuticle of Ptilophyllum leaves are hypostomatic condition with syndetocheilic stomata, but vary in the pattern of distribution of stomata, papillae and its morphological differences, being the main criteria for speciation. P. acutifolium Morris has pinnae with acute apices, whereas P. cutchense Morris possesses sub-acute to obtuse apices. P. rarinervis (Feistmantel) Bose has pinnae with 2-3 veins only. P. tenerrimum Feistmantel has narrow, long pinnae. In P. indicum Jacob & Jacob 1954, stomata are distributed in stomatal bands. P. oldhami Jacob & Jacob 1954 shows irregular distribution of stomata except on the marginal region. In P. distans Jacob & Jacob 1954 pinnae have acute apices and the lower cuticle is differentiated into broad stomatiferous and narrow non-stomatiferous bands, cells are devoid of papillae and the outer wall of the subsidiary cells is looped. P. jabalpurensis Jacob & Jacob 1954 is characterised in having 1-3 papillate marginal cells and papillae are solid circular or semi-circular, hollow-circular or crescent-shaped. In P. nipanica Vishnu-Mittre 1956 lower epidermis has non-papillate cells and stomata are arranged in bands. P. gladiatum Bose & Sukh-Dev 1958 is distinct in having gladius-like pinnae and nonpapillate epidermal cells of veins and subsidiary cells. P. institacallum Bose 1959 is significant in possessing papillae of various shapes. The epidermal cells of lower cuticle and stomata are irregularly scattered in the inner zone. P. sakrigaliensis Sah 1958 has elongated pinnae with acute apices. Lower cuticle is studded with stomata arranged in bands. One or two papillae are present on each epidermal cell of the vein band. P. borridum Roy 1967 shows lower cuticle with stomata in bands and two or more papillae on each epidermal cell of the vein band but they do not form a frill-like structure over the vein band as in P. institacallum. P. sabnii Gupta & Sharma 1968 has small pinnae, oblong or oval in shape with obtuse apices, lower cuticle is differentiated into broader stomatal, narrow nonstomatal band and marginal non-stomatal region, epidermal and subsidiary cells are devoid of papillae. P. amarjolense Bose 1953b is distinguished by its broader leaves and stomatal bands are 4-5 stomata broad.

#### Male flower-Weltrichia Braun

The male flower is characterised by having microsporophylls forming a cup-shaped structure at



Text-figure 3-A, Ptilopbyllum-pinna showing venation; B, Ptilopbyllum leaf; C, T.S. of pinna; D, T.S. of rachis.

the base. They bear polleniferous appendages adaxially or distally. Each appendage has two rows of pollen chambers. The pollen grains are oval and monocolpate.

From India, male flower was first described as *Williamsonia santalensis* Sitholey & Bose 1953. Later, Bose (1967) suggested that the specimen of *Williamsonia santalensis* be included under the genus *Weltrichia* Braun as it has priority. The male flower consists of a whorl of 20 (?)microsporophylls coalescent towards the base to form a shallow cup. Each microsporophyll bears on its upper surface two rows of finger-like appendages which are presumed to have been the spore-bearing organs. The rows of small chambers are present inside each appendage. The distal sterile portion of the microsporophyll shows an asymmetrical and obliquely twisted

expansion of the lamina. Sharma (1969a) suggested that there are two whorls, the outer whorl consists of 20 sterile bracts and the inner whorl has 20 twisted microsporophylls. He also opines that the appendages are in two rows on the adaxial side of the proximal part of the microsporophyll and there is only one row on the distal part of the microsporophyll (Text-figure 4C). Later, Sitholey and Bose (1971) commented that the bracts are spirally arranged and are not in whorls. The receptacle of Weltrichia is, in fact, cup-shaped flower formed by the fusion of microsporophylls. The fertile organ consists solely of a tubular thickwalled structure, tapering distally which completely encloses the polleniferous chambers. Revised restoration of microsporophyll given by Sitholey and Bose (1971) shows that the microsporophylls were

bilaterally symmetrical structures without any twisting of distal portion. The abaxial surface of the expanded distal portion is raised into a prominent hump-like structure (Text-figure 4A, B).

Ontheanthus polyandra Ganju 1947, a unique bennettitalean fructification, was transferred by Sharma (1969a) to Williamsonia santalensis Sitholey & Bose 1971 recognised it as a separate taxon under Weltrichia. It needs further examination whether it is a part of an unopened flower of Weltrichia santalensis or an incomplete preserved part of a bisexual flower-Amarjolia dactylota. Weltrichia singhii Bose 1967 is supposed to represent bud conditions of W. santalensis (Sharma, 1969a). Williamsonia companulatiformis, known by an incomplete specimen, is built more or less on the same plan of construction as Weltrichia santalensis (Sharma 1969a, p. 102; Sitholey & Bose 1971). Weltrichia harrisiana (Bose & Banerji, 1984) is about 12-15 cm in diameter. Microsporophylls 12-14 in number, coalesce towards the base and form a circular depression, about 2 cm in diameter. The microsporophylls are distally expanded, coriaceous and their adaxial surface exhibits prominent ridges. Each microsporophyll terminates into five uniformly broad fertile appendages, each appendage having a double row of polleniferous chambers. Weltrichia maldaensis Pal & Ghosh 1985 shows microsporophylls with some small circular lateral projections.

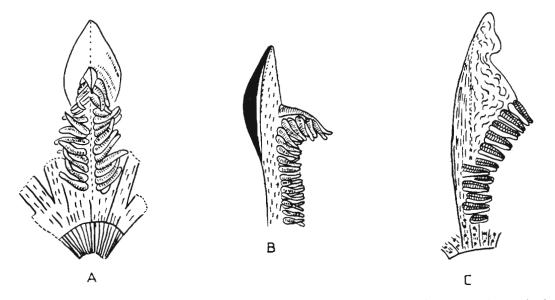
#### Female flower-Williamsonia Carruthers

*Williamsonia* Carruthers, the female flower, has receptacles of various shapes, e.g., rhomboidal,

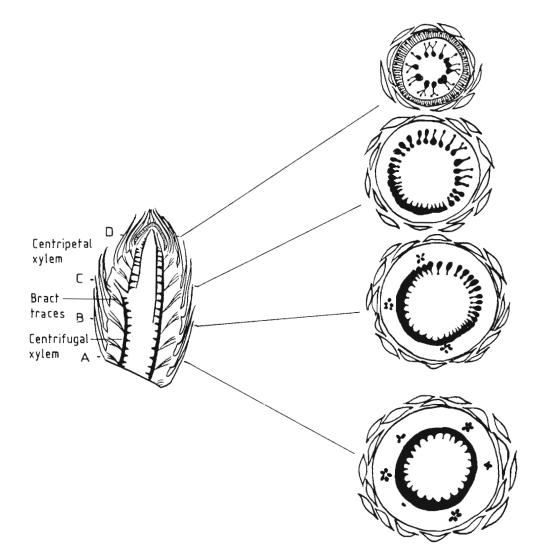
hemispherical-conical with numerous interseminal and seminal scales. The receptacle is surrounded by numerous spirally arranged bracts.

The anatomy of peduncle (Text-figure 5) shows the changes in the vascular organisation at different levels (Sharma, 1973b). The centrifugal xylem is gradually reduced in thickness and broken into segments. Centripetal xylem originates from the endarch protoxylem points of centrifugal xylem and gradually increases in amount towards the upper side of peduncle. In the receptacle centripetal xylem forms the main vascular cylinder and the remnants of the centrifugal xylem are connected with each other by their tangentially elongated protoxylem points.

Anatomy of receptacle shows the main stele consisting of a number of isolated, collateral inverted, exarch vascular bundles which maintain their shape and size from base to apex (Text-figure 6). In the basal position bundles gives off paired traces to the bracts, while in the middle part of the receptacle they are connected with isolated, collateral or concentric, endarch or mesarch vascular bundles present in the peripheral region of cortex. These peripheral bundles are the source of vascular traces to the scales. The tracheids of bract traces have spiral thickening on their lateral walls. Transverse section through the concavity of receptacle shows inverted bundles in the main stele indicating a change from endarch condition of vascular bundle in the peduncle to exarch condition in the receptacle. The main stele gives off traces only at two places in the whole receptale. The first point lies in the pedicel while the second is present in the middle part of receptacle. These two points in the



Text-figure 4-Restoration of microsporophyll-A, B, after Sitholey and Bose (1971); C, after Sharma (1969a).



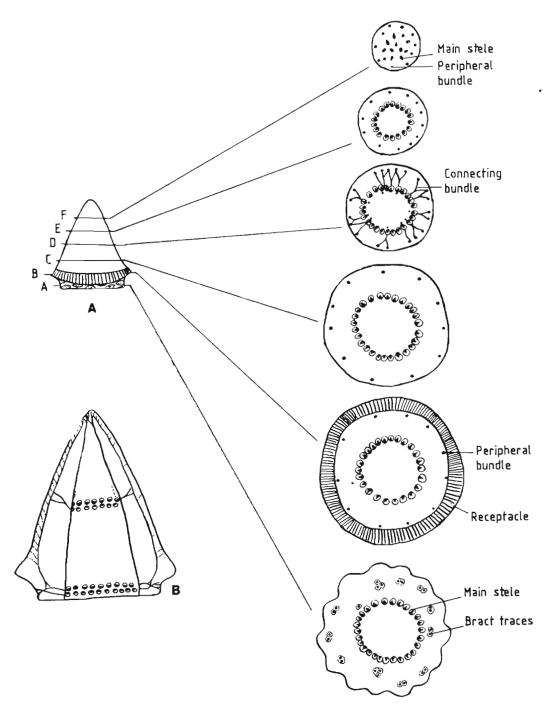
Text-figure 5-Reconstruction of the vascular organisation in transverse and longitudinal section of peduncle.

receptacle may be compared with the two modes of a branch or stem. According to Sharma (1973b) the receptacle of a seed-bearing *Williamsonia* is comparable with a two noded inflorescence axis.

Bract anatomy shows a thick sclerenchymatous hypodermal zone which consists of relatively larger cells on the abaxial side. There are usually 7 collateral, endarch vascular bundles, two above near the middle line and five forming an arc parallel to the lower surface. The xylem of the bundles facing each other in the radial portion of the bract and in the distal flattened part, the number of bundles reaches up to 11 or more. The ground tissue is parenchymatous but without mucilage canals (Sahni, 1932; Sharma, 1975).

The upper surface of the bracts has rectangular or squarish epidermal cells which are arranged in rows, parallel to the long axis of the bracts. The lateral walls of the cells are thin and sinuous. Outer wall of the epidermal cells is provided with circular, scattered simple pits. Stomata are syndetocheilic, non-sunken and transversely oriented.

The fertile portion consists of a small, conical receptacle which is produced terminally on a thick peduncle. The receptacle is covered with a thick layer of sterile and fertile scales. The sterile scales consist of hexagonal-rectangular parenchymatous cells with a central vascular strand up to the highly cutinized distal regions. Five or six sterile scales surround a fertile seminiferous scale represented by an orthotropous ovule which consists of a small, thick stalk or funiculus and a long, oblong body. The stalk has a definite concavity at its basal end, representing the abscission point of the scale. From the upper part of funiculus arise a central long, slender nucellar stalk and a thin outer integument. The former consists of two types of cells, central thin-walled parenchymatous and the surrounding



**Text-figure 6–A**, Transverse section of the receptacle showing vascular organisation; **B**, Longitudinal section of the receptacle showing vascular organisation.

thick-walled, darkish, elongated cells. The nucellar stalk terminates into an oval-shaped nucellus, the apical end of which is made up of a rosette of elongated cells. The nucellus is fused with integument which is made up of rectangular, parenchymatous cells. It is three or four cells thick in the region of micropyle but reduces downward, so that near the basal part of nucellus it is only one cell thick and then merges into the surrounding layer of nucellar stalk.

The distal end of micropyle is more or less a funnel-shaped structure and reaches the level of the terminal end of the surrounding interseminal scales. The outer integument arises from the funiculus remains which are thin, delicate and mostly one cell thick throughout its length. It remains free from the

**Text-figure** 7—Vertical section of ovule—**A**, after Sahni (1932), and **B**, after Sharma (1975).

nucellar stalk, but fuses with the inner integument near the basal part of the nucellus and then continues upwards as an outermost layer of integument up to the distal end of micropyle (Textfigure 7A, B.

The different developmental stages of an ovule show that the development of fertile layer is basipetal (Sharma, 1974b). In the basal part of receptacle the fertile layer consists of thin-walled polygonal cells without any distinction of scales. The scales are produced as a result of ingrooving of epidermis. Differentiation of fertile scale starts from the micropylar end. The development of integument takes place at an early stage of ovule ontogeny. In a young ovule there is a long cylindrical nucellar stalk which terminates into a conical nucellus; pollen chamber is differentiated in the upper part of nucellus (Text-figure 8A). A megaspore mother cell is produced in the nucellus which divides by free nuclear divisions for the formation of an endosperm. The endosperm wall formation starts from the micropylar side and continues on the descending side (Text-figure 8B). On maturity of endosperm the surrounding cells of nucellus degenerate, except the micropylar side (Text-figure 8C). Pollen chamber formed by the degeneration of nucellar cells shows affinity of Williamsonia with the cycads on one hand and Ephedra on the other (Sharma, 1974b). Sharma (1979) further observed that the megaspore mother cell is situated not very deep in the nucellus. The female gametophyte covers almost whole nucellus and is made up of thin-walled cells. In the apical part of endosperm two or more archegonia are seen

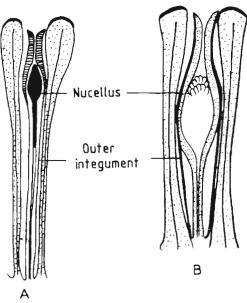
arranged in a ring and distinct from the cells of endosperm. The micropylar canal in young ovule is long, narrow and lined with large cells, while in mature ovules or seeds the canal becomes small, wide and filled by a nucellar plug. The thickness of micropyle reduces from 3-4 cells to one showing relationship with Cycadeoides and Gnetum. Mature seeds are found in ripe, nacked, bractless fruits. Seeds are oblong or ovate, stalked, dicotyledonous, partially endospermic and have a long micropyle (Sharma, 1970c). Seed coat includes outer tubular cells, middle parenchymatous true integument and inner vascularized tissue formed after fertilization by modification of the peripheral portion of nucellus. The seed of Williamsonia differs from cycadeoideas in being partially endospermic and shows more affinities with the seeds of pteridosperms and Gnetales than with the angiosperms.

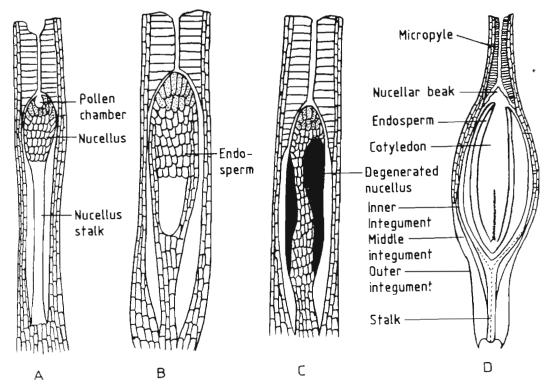
On maturity, the flower produces abscission layer in between the receptacle and the compact layer of sterile and fertile scales. The sterile scales and ovules are ontogenetically identical. It is related to the extant genus *Ginkgo* in the manner of detachment of flower from the parent plant before fertilization. The inter-locking cells of the micropyle was either absent or the flower was detached before pollination.

## ATTACHMENT OF FLOWER

On the basis of their plan of construction the flowers can be divided into two main types, viz., "exposed type" as in W. guptae and W. amarjolense Sharma and partially "hidden type" as in W. sewardiana Sahni. In exposed type, the flower were borne either terminally on the branches or directly on the sides of the stem. They had a definite point of abscission and covered with bracts produced from the base of the receptacle. Moreover, on the basis of number, shape and size of bracts they can be further divided into three main types (Sharma, 1971). In the first type, the bracts are large, incurved and mostly arranged in one row as in W. amarjolense (Textfigure 9A). The second type has long, linear bracts arranged in two rows as in W. guptae (Text-figure 9B). The third type includes the flower with linear, thin, straight and arranged in several rows of bracts as in W. cf. W. scotica (Text-figure 9C). The second type of partially embedded flower probably detached itself from the parent plant on maturation leaving a depression on the side of the stem (Textfigure 9D). The variation on the basis of number, shape and size of bracts might be due to seasonal fluctuation.

Feistmantel (1876) described Williamsonia





Text-figure 8-A-C, Vertical section of ovules showing different stages of development; D, Seed restoration.

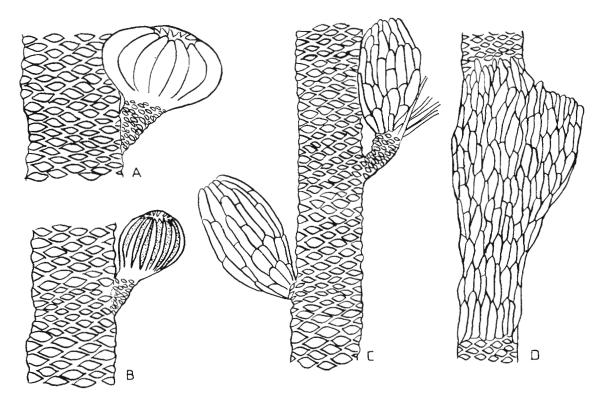
blanfordi from Kachchh. W. gigas Carruthers recorded by Feistmantel (1877a) has been ultimately placed under Weltrichia santalensis. W. microps Feistmantel 1877a is visualized as a small pedicellate bud like specimen of W. gigas or W. blanfordi by Seward (1917, p. 445) and Seward and Sahni (1920). Williamsonia comb. W. gigas Feistmantel 1877b has been placed under a new species-W. indica, by Seward (1917) in being larger in size. In W. sewardiana (Sahni, 1932) the flower has hemispherical receptacle, seminal and interseminal scales 3 mm in length which gradually reduce up to 1.5 mm. Soft investment around the nucellus continued downwards forming the outer layer of nucellar stalk. Sharma (1975) observed adherence of nucellus and integument throughout its length (Text-figure 7B). W. sahnii Gupta 1943 is supposed to be a bisexual flower but as there is no positive evidence of the presence of androecium, the author himself doubts about its bisexuality. W. harrisiana Bose 1968 is characterised by dome-shaped receptacle, seminal and interseminal scales up to 1 cm in length and the presence of a sterile disc. Sharma (1977) observed sterile disc in almost all the species.

*W. guptae* Sharma 1968 has conical receptacle with seminal and interseminal scales up to 1 cm in length, epidermal cells of the bract rectangular and stomata sunken. *W. amarjolense* Sharma 1968 is

distinct in having hemispherical receptacle, epidermal cells of bracts are polygonal and stomata non-sunken. Perhaps W. guptae is similar to W. *harrisiana* in shape of receptacle, size of seminal and inter-seminal scales and epidermal features of the bracts. W. seniana Bose & Kasat 1969 can be distinguished by its epidermal features. In W. cf. W. scotica, the bracts are considered as reduced pinnate leaves having pinnae in the form of reduced vascularised appendages (Sharma, 1970a). W. kakadbhitensis Bose & Banerji 1984 is distinct in having rhomboidal receptacle and hairy bracts. W. trambauensis (Bose & Banerji, 1984) is characterized in having ramentum scars and trichome bases. W. sukhpurensis (Bose & Banerji, 1984) has jagged-thickening on the lateral walls of epidermal cells.

### HABIT OF WILLIAMSONIA PLANT

On the basis of morphological and anatomical characters, it is suggested that the plants of *Williamsonia* were perhaps small, much branched woody trees of xerophytic environment. They had adopted various ecological strategies to meet the water stress conditions as shown by different parts of the plants. The columnar stem was branched monopodially or sympodially, bearing large and small slender branches with spirally arranged



Text-figure 9-A, B, Showing attachment of flower on lateral branch; C, Reconstruction showing terminal or lateral attachment as shown by *Williamsonia* cf. *W. scotica*; D, Partially hidden type of *Williamsonia* flower.

pinnate foliage leaves. The pedunculate flowers were borne terminally/laterally at the point of dichotomy or the sessile flowers were partially embedded into the stem. These woody trees perhaps formed a part of mixed subtropical community of the forest. The palaeolatitudinal position of the Indian subcontinent during the period was between 20° 45° south latitude (Smith & Briden, 1979) and this supports the existence of tropical to subtropical climate. The stem of Williamsonia possesses persistent armours of leaf-bases which perhaps acted as a protective covering either from predators or to withstand unfavourable conditions. The armours might have provided mechanical strength to the stem also. The variation in size and shape of the leaf scars depends upon the position of abscission layer and is probably environmentally induced variation. The presence of periderm and the stem anatomy show arborescent nature of the plant. The periderm protects the inner tissue from weathering. The presence of sclereids in the peduncle and rachis, sclerenchyma cells outside and in between the vascular bundle of the rachis provides hardness to these parts. Stem has a large pith and less secondary tissue suggesting that the plant could grow in moisture deficit soil. The secondary wood shows large lumened variously pitted tracheids in the growing season (Early wood) indicating that water

availability was high. Thick-walled tracheids with less variable pitting (Late wood) show water stress condition. The leaf anatomy shows sheathed vascular bundles. Sclerenchymatous hypodermis acts as a supporting tissue and provides rigidity to the leaves to withstand wilting. The occurrence of sunken stomata on the lower surface (hypostomatic condition) in most of the species and sclerenchymatous hypodermis show xeromorphic characters. Sclerenchyma and sclereids give mechanical strength and protection. Papillae in the epidermal cells as well as subsidiary cells act in two ways, i.e., they reduce the intensity of the incoming radiation and also check the wind action. High stomatal frequency of occurrence in a few species perhaps is due to high rate of transpiration. Growth rings in the secondary wood and abscission of leaf show seasonal fluctuation in light, temperature and water availability. The mechanism of abscission gives protection against desiccation in adverse conditions. Abscission of bracts shows maturity of the flowers. The abscission of the reproductive parts, i.e., seminal, interseminal scales and microsporophylls from the receptacle base helps in dissemination for fertilization and manner of dispersal. Hairs on the bracts perhaps protect the reproductive parts from insect predators and also reduce the transpiration rate. The ovules are further

protected by interseminal scales which possibly acted as protective covering but, later attracted the insects for pollination. In *Williamsonia*, flowers were unisexual suggesting out-breeding by anemophilly, whereas *Amarjolia*-flower (another bennettitalean flower) was hermaphrodite. The curved nature of microsporophyll and protandrous nature of *Amarjolia* also indicate out-breeding by entomophily. Whether these major selective characters provide protection from phytophagus insects or from self-incompatibility are questions to be considered squarely.

A cycas-like habit for Williamsonia sewardiana was proposed by Sahni (1932), plant with columnar stem-Bucklandia indica Seward, crown of pinnate foliage leaves-Ptilophyllum cf. cutchense Morris and female flower borne terminally on short branched laterals-Williamsonia sewardiana. Later, the recovery of more species of female flower: W. guptae Sharma, W. amarjolense Sharma, Stem-Bucklandia sahnii Bose, and leaves-Ptilophyllum amarjolense Bose and P. sahnii Gupta & Sharma from the same locality raised doubts about Sahni's reconstruction of W. sewardiana, Bucklandia indica and Ptilophyllum cf. cutchense. Anatomically the peduncle of W. sewardiana is more similar to Bucklandia sahnii Bose than with B. indica Seward (Bose, 1953a; Sharma 1975). Besides, the discovery of branched stem of B. sahnii and B. dichotoma suggested that these plants had equal or unequal

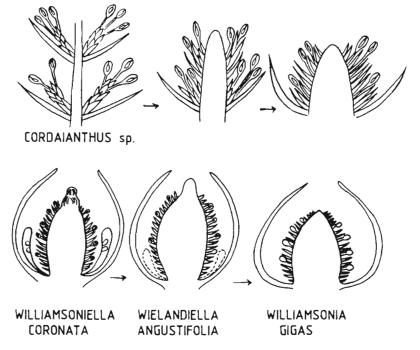
branched stems and perhaps due to the deciduous nature of small laterals and leaves they were found in large number. Recently, a specimen has been recovered from Rajmahal Hills in which the *Williamsonia*-flower seems to be laterally attached to a narrow branch perhaps by a small peduncle. On the basis of these evidence, the branching in *Williamsonia* was probably more akin to the reconstruction of Harris (1969, fig. 59c) for *Bucklandia pustulosa, Ptilopbyllum pecten* and *Williamsonia leckenbyi*. This view has also been supported by Bose (1974, p. 197), but it needs better specimens with branching habit.

# **RELATIONSHIPS OF WILLIAMSONIA**

*Williamsonia*, belonging to Bennettitales, shows primitive as well as advance gymnospermous characters. It shows affinities with the plants of Pteridospermales, Cordaitales, Cycadales, Gnetales, etc.

## Affinities with Pteridospermales

Presence of secretory canals in pith and cortex of stem shows affinity with the Pteridospermales. In the petiole the bundles are united at places exhibiting a continuous C-shaped strand as in some pteridosperms, viz., *Rhetinangium arberi* Gorden. The endospermic seed of *Williamsonia* also shows



**Text-figure 10**-Stages showing derivation of an ovulate cone of *Williamsonia* from a female inflorescence of *Cordaiantbus* sp. as a result of condensation and swelling of the axis.

affinity with the seed of pteridosperms. The microsporangiate flower having pinnate microsporophylls shows some similarities with pteridosperms. But, in *Weltrichia*, a cup-shaped receptacle with microsporophylls and microsporangia which are either directly attached or arranged on finger-like appendages are those notable characters not exhibited by an extinct or extant group of plants.

# Affinities with Cordaitales

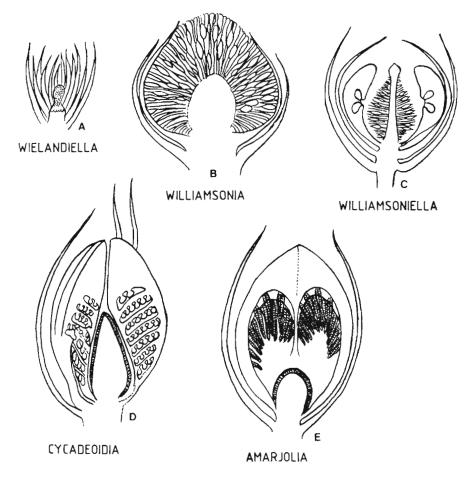
Sharma (1982) derived bennettitalean seed bearing fructification from *Cordaianthus* as a result of reduction and swelling of inflorescence axis producing receptacle and modification of the axillary bud bearing megasporophylls and scales to seminiferous and interseminal scales (Text-figure 10). However, in ovule ontogeny *Cordainthus* and *Williamsonia* are different from each other.

# Affinities with Cycadales

Presence of armours of leaf bases on the stem, crown of pinnate leaves and secretory canals in pith and cortex shows similarity with that of *Cycas*. Girdle leaf traces are absent unlike *Cycas*. Cuticle and anatomy of leaf also differ from *Cycas*. In *Williamsonia*, stomata are syndetocheilic and transversely oriented, whereas in Cycadales stomata are haplocheilic. Diploxylic condition in the anatomy of *Cycas* leaf is also unlike Williamsonias. In *Cycas*, petiole shows omega-shaped, collateral endarch bundles, whereas in *Williamsonia* they are arranged in double row. Non-vascularised integument of ovule differs from cycads but the megagametophyte with 3-4 archegonia and pollen chamber formed by the degeneration of nucellar cells are the common features.

# Other affinities

Mehra (1988) derived Bennettitales independently from Protoseminales, a plexus which gave rise to Cordaitales and Pteridospermales. Presence of definite abscission point at the base of fructification suggests similarity with the *Ginkgo*. In *Williamsonia*, the integument surrounding each

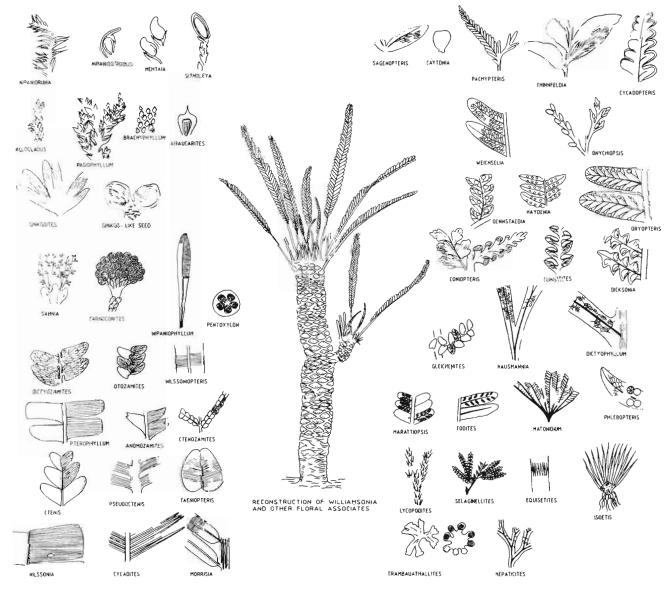


Text-figure 11—Showing gradual sequence of flower from monosporangiate to bisporangiate condition or vice versa; A, Weilandiella;
B, Williamsonia; C, Williamsoniella; D, Cycadeoidea; and E, Amarjolia.

ovule and extending up to a long micropyle exhibits an advanced feature shown by *Gnetum* and *Welwitschia*. The formation of pollen chamber by the degeneration of nucellar cells shows its relationship with *Ephedra*. Flower-like organisation of *Williamsonia* fructifications led Wieland (1906) and Arber and Parkin (1907) to suggest that Mesozoic Bennettitales are the ancestral group of angiosperms. A gradual sequence can be drawn from monosporangiate to bisporangiate flower, i.e., *Wiellandiella, Williamsonia, Williamsoniella, Amarjolia* to *Cycadeoidea* or vice versa (Text-figure 11), but which one is primitive is difficult to say.

# TIME OF WILLIAMSONIAS

The Williamsonias appeared in India during Upper Jurassic Period and flourished well in tropical to subtropical climate of Lower Cretaceous Period. The other floral associates of Williamsonias are represented by extremely variable taxa of highly diversified plant groups (Text-figure 12). They are: Hepaticites Walton, Trambauathallites' Banerji, Equisetites Sternberg, Lycopodites Lindley & Hutton, Isoetes Linnaeus, Selaginellites Zeiller, Marattiopsis (Oldham & Morris) Seward & Sahni, Todites Seward, Klukia Raciborski, Gleichenites Goeppert, Matonidium Schenk, Phlebopteris Brongniart, Hausmannia Dunker, Dictyophyllum Lindley & Hutton, Haydenia Seward, Dicksonia L'Heritier, Culcitites Appert, Onychiopsis Yokoyama, Coniopteris Brongniart, Dennstaedtia Bernhardi, Weichselia Stiehler, Pachypteris Brongniart, Cycadopteris Zigno, Thinnfeldia Ettingshausen, Sagenopteris Presl, Caytonia Thomas, Pterophyllum Brongniart,



Text-figure 12-Reconstruction of Williamsonia (after Sahni, 1932) and other floral associates.

Otozamites Braun, Dictyozamites Oldham, Anomozamites Schimper, Nilssoniopteris Nathorst, Pentoxylon Srivastava, Nipaniophyllum Sahni, Carnoconites Srivastava, Sahnia Vishnu-Mittre, Nilssonia Brongniart, Morrisia Bose, Pseudoctenis Lindley & Hutton, Ctenozamites Nathorst, Cycadites Sternberg, Taeniopteris Brongniart, Ginkgoites Seward, Araucarites Presl, Brachyphyllum Lindley & Hutton, Pagiophyllum Heer, Nipanioruha Rao, Elatocladus Halle, Nipaniostrobus Rao, Mehtaia Vishnu-Mittre, Sitholeya Vishnu-Mittre, Podostrobus Rao & Bose and Stachyotaxus Nathorst.

The Williamsonias were one of the dominant plant groups growing in dry open lands alongwith pteridophytes and conifers and perhaps formed a part of tropical semi-evergreen and subtropical broad-leaved type of forest. In India, they appeared and established themselves during the Upper Jurassic (marine), proliferated in the Lower Cretaceous and disappeared in the Upper Cretaceous with the advent of the highest evolved plant group the angiosperms perhaps due to inherent genetic factors. The rapid radiation of angiosperms caused the sudden extinction of Williamsonias due to strong selective pressure. Perhaps they could not compete with the early angiosperms and ultimately perished from the Upper Cretaceous floristic scenario.

#### REFERENCES

- Arber EAN & Parkin J 1907. On the origin of angiosperms. Bot. J. Linn. Soc. 38: 239.
- Bose MN 1953a. Bucklandia sabnii sp. nov. from the Jurassic of the Rajmahal Hills, Bihar. Palaeobotanist **2** : 41-50.
- Bose MN 1953b. Ptilophyllum amarjolense sp. nov. from the Rajmahal Hills, Bihar. Proc. natn. Inst. Sci. India 19(5): 605-612.
- Bose MN 1959. The fossil flora of the Jabalpur group 1-Ptilophyllum institacallum n. sp. Palaeobotanist 7: 26-28.
- Bose MN 1967. Weltrichia singhii n. sp. from the Rajmahal Hills, Bihar. Curr. Sci. **36**(2): 48-49.
- Bose MN 1968. A new species of *Williamsonia* from the Rajmahal Hills, India. J. Linn. Soc. (Bot.) **61**(384) : 121-127.
- Bose MN 1974. Bennettitales. In Surange KR, Iakhanpal RN & Bharadwaj DC (editors)—Aspects & Appraisal of Indian Palaeobotany: 189-200. Birbal Sahni Institute of Palaeobotany, Lucknow.
- Bose MN & Banerji J 1984. The fossil flora of Kachchh. I-Mesozoic megafossils. *Palaeobotanist* **33** : 1-189.
- Bose MN & Kasat ML 1969. The fossil flora of the Jabalpur Series : 4. Williamsonia seniana n. sp. In Santapau H et al. (editors)—J. Sen Mem. Vol. : 305-309. Bot. Soc. Bengal, Calcutta.
- Bose MN & Sukh-Dev 1958. A new species of *Ptilopbyllum* from Bansa, south Rewa Gondwana Basin. *Palaeobotanist* **6** : 12-15.
- Feistmantel O 1876. Jurassic (Oolitic) flora of Kach. In Fossil flora of the Gondwana System. Mem. geol. Surv. India Palaeont. indica ser II, 2(1): 1-80.

Feistmantel O 1977a. Jurassic (Liassic) flora of the Rajmahal

Group in the Rajmahal Hills. In Fossil flora of the Gondwana System. Mem. geol. Surv. India Palaeont. indica ser. II,  $1(2) : 1 \cdot 10$ .

- Feistmantel O 1877b. Jurassic (Liassic) flora of the Rajmahal Group from Golapili (Near Ellore), South Godavari District. Mem. geol. Surv. India Palaeont. indica, Ser II, 1(3): 1-28.
- Ganju PN 1947 Ontheanthus polyandra gen. et sp. nov., a new type of gymnosperm male fructification from the Rajmahal Hills. Proc. Indian Acad. Sci. **B25**(5): 105-118.
- Grant CW 1840. Memoir to illustrate a geological map of Cutch. Trans. geol. Soc. ser 2, 5(2): 289-329.
- Gupta KM 1943. A new species of *Williamsonia* from the Rajmahal Hills. J. Indian bot. Soc. **22** : 191-199.
- Gupta KM & Sharma BD 1968. Investigations on the Jurassic flora of the Rajmahal Hills, India 2. On a new species of *Ptilophyllum*, *P. sabnii* from Amarjola in Amrapara region. *J. palaeont. Soc.* 11: 1-7.
- Harris TM 1969. The Yorkshire Jurassic flora III-Bennettitales. Br. Mus. nat. Hist., Lond.: 1-186.
- Jacob K & Jacob C 1954. Cuticular study of India: *Ptilopbylli*<sup>499</sup>. fronds from Cutch and Jubbulpore. *Mem. geol. Surv. India Palaeont. indica* **33**(1): 1-35.
- Lignier O 1908. Le fruit de Bennettites et l'ascendance de Angiospermes. *Bull. bot. Soc. France* **55** : 1-17.
- Mehra PN 1988. Indian conifers, gnetophytes and phylogeny of gymnosperms. Bot. Department, Panjab Univ., Chandigarh.
- Pal AK & Ghosh SC 1985. Weltrichia (male Williamsonia) from the bore-hole samples of Malda, West Bengal, India. Curr. Sci. 54(12): 574-576.
- Roy SK 1967. Ptilopbyllum horridum sp. nov. from Trambau, Kutch. Curr. Sci. 36(21): 581-582.
- Sah SCD 1958. Ptilophyllum sakrigaliensis n. sp. from Sakrigalighat, Rajmahal Hills, Bihar. Proc. 45th Indian Sci. Congr., Madras 3: 337.
- Sahni B 1932. A petrified *Williamsonia* (*W. sewardiana* sp. nov.) from the Rajmahal Hills. *Mem. geol. Surv. India Palaeont. indica* 20: 1-19.
- Seward AC 1912. A petrified Williamsonia from Scotland. Phil. Trans. R. Soc. 203 : 101-126.
- Seward AC 1917 Fossil plants III. Cambridge.
- Seward AC & Sahni B 1920. Indian Gondwana plants: a revision. Mem. geol. Surv. India Palaeont. indica 7(1): 1-41.
- Sharma BD 1967. Investigations on the Jurassic flora 4. On a new species of Indian *Bucklandia*, *B. guptai* with remarks on *B. sahnii* Bose of Rajmahal Hills, India. *Ameghiniana* 4(2): 35-46.
- Sharma BD 1968. Epidermal studies on the bracts in two new species of Williamsonia, W. guptai and W. amarjolense. Acta bot. Hung. 14 : 373-383.
- Sharma BD 1969a. Further observation on Williamsonia santalensis Sith. & Bose with description of a new species. Palaeontographica B125(4-6): 93-103.
- Sharma BD 1969b. Bucklandia dichotoma sp. nov. from the Middle Jurassic of Rajmahal Hills, India. Ameghiniana 6(4): 303-308.
- Sharma BD 1970a. On the structure of Williamsonia cf. W. scotica from the Middle Jurassic rocks of the Rajmahal Hills, India. Ann. Bot. 34(135): 289-296.
- Sharma BD 1970b. On the vascular organisation of the receptacle of seed-bearing Williamsonias from the Middle Jurassic rocks of Amarjola in the Rajmahal Hills, India. Ann. Bot. 34(138) : 1063-1070.
- Sharma BD 1970c. On the structure of the seed of *Williamsonia* collected from the Middle Jurassic rocks of Amarjola in the Rajmahal Hills, India. *Ann. Bot.* **34**(138) : 1071-1077.
- Sharma BD 1971. On a collection of bennettitalean stems and

fructification from Amarjola in the Rajmahal Hills, India. *Palaeontographica* **133**(1·3): 48-52.

- Sharma BD 1973. Anatomy of the peduncle of *Williamsonia* collected from the Jurassic of Amarjola in the Rajmahal Hills, India. *Botanique* 4(2): 93.100.
- Sharma BD 1974. Pollen chamber in the ovule of Williamsonia Carr. Curr. Sci. 43(1): 22.
- Sharma BD 1975. Observation on Williamsonia sewardiana Sahni from Amarjola, Rajmahal Hills, India. Cas. Miner. Geol. 20(1): 31-37.
- Sharma BD 1977. Indian Williamsonias—an illustrated review. Acta palaeobot. 18(1): 19-29.
- Sharma BD 1979. Archegonia in *Williamsonia* Carr. (Bennettitales). *Curr. Sci.* **48**(13) : 601.

Sharma BD 1982. Morphology of interseminal scales in William-

sonia Carr. (Bennettitales). Indian J. Earth Sci. 9(1): 1-6.

- Sitholey RV & Bose MN 1953. Williamsonia santalensis sp. nov. a male fructification from the Rajmahal Series with remarks on the structure of Ontheanthus polyandra Ganju. Palaeobotanist 2: 29-39.
- Sitholey RV & Bose MN 1971. Weltrichia santalensis (Sith. & Bose) and other bennettitalean male fructifications from India. Palaeontographica B131(5-6): 151-159.
- Smith AG & Briden JC 1979. Mesozoic and Cenozoic palaeocontinental maps. Cambridge University Press, Cambridge, London, New York, Melbourne.
- Vishnu-Mittre 1956. Studies of the fossil flora of Nipania, Rajmahal Series, India—Bennettitales. *Palaeobotanist* **5** : 95-99.
- Wieland GR 1906. American fossil cycads-I. Carneige Institute, Washington.